



Summary of Health Data for DOW CORNING™ C6-265 Elastomer

SUMMARY

Cured DOW CORNING™ C6-265 Elastomer was tested to determine potential to cause dermal sensitization, and cytotoxicity, as well as local and systemic effects in a USP Class V test and a short-term implant test. When tested directly or by extract exposure, the material was not a skin sensitizer. The material passed the systemic and intracutaneous reactivity tests that compose the USP Class V test and also met the requirements of 30-day implant study (with interim sacrifice period) in rabbits.

Note: The toxicology information in this data sheet is limited to that which is specified to meet C6 standards. Additional toxicology information, if available, may be provided to you upon request for an additional charge.

TOXICOLOGY DATA

Acute Toxicity

Skin Sensitization. The cured test article was tested for skin sensitization potential in guinea pigs in a modified Buehler test. Guinea pigs were exposed to the test article (solid), or extracts of the test article prepared in 0.9% Sodium Chloride for Injection (SCI), or acetone. The extracts were prepared at a ratio of 3 cm² test article per ml of extractant. The SCI extracts were prepared by heating at 121±2 °C for 60 minutes in an autoclave, and the acetone extracts were prepared by heating at 70±2 °C for 24 hours in an incubator. An irritation screen in 2 guinea pigs was conducted to determine the irritation potential of the solid test article, the SCI and acetone extracts (100%) of the test article, and the extraction medium (acetone). The screen involved a 6-hour exposure to test and control materials, followed by an evaluation for erythema and edema at 24 and 48 hours post-treatment. No irritation was observed in either animal at sites with any treatment. In the main study, the 10 animals in each test group were exposed to the solid (1 cm²), or extracts (SCI and acetone, 100%, 0.5 ml under a 25 mm Hilltop chamber). All animals were wrapped with occlusive dressings for 6 hours, once per week, during the 3-week induction phase. Scores were determined as in the screening test. Fourteen days after the final induction, the test animals were challenged with the same treatments as during the induction phase for 6 hours at naïve sites. Five animals were employed in the control group. Each of the control animals was induced with acetone alone, then challenged with the test article (solid), SCI extract (100%), and acetone extract (100%) in a similar manner. In addition, a positive control study was conducted to validate the sensitivity of the



test system and alpha-hexylcinnamaldehyde (50% and 3% in PEG 300, respectively, for the induction and challenge phases) was selected as the positive control material. Dermal observations for sensitization were recorded at approximately 24 and 48 hours after challenge patch application. There were no scores noted for any of the test animals. One animal of the Acetone Extract Test group died on the 24-hour reading after the third induction (Day 16). The cause of death was considered to be spontaneous and unrelated to treatment because no macroscopic finding was identified. The positive and negative controls performed as expected. Under the conditions of this study, the test article and its extracts in saline or acetone did not exhibit the potential to produce dermal sensitization in guinea pigs (1).

Biocompatibility

Cell Culture. The cytotoxic effects of the cured test article were determined with cells in culture. Duplicate 1 cm² samples of test material were each placed in a cell culture well containing MRC-5, human embryonic lung cells. The materials were placed in direct contact with the cell monolayer and incubated on the cultures for approximately 24 hours at 37°C. Separate samples of the test material were extracted for 24 hours at 37°C at a surface area to volume ratio of 3 cm² per milliliter of Minimal Essential Medium (MEM). To test the extract, the growth medium was removed from the cultures and replaced with an extract; the cells were then incubated for approximately 48 hours at 37°C. A positive control for the direct cell contact and extracts (reference standard of the British Standards Institute) and a negative control (SRM-C, a high-density polyethylene) were employed in the test system in the same manner. After incubation, the cells received a morphological evaluation using a light microscope. In addition, the cells exposed to the extract were tested for their ability to take up a vital dye, Neutral Red. Sodium lauryl sulfate served as the positive control for cell viability testing. No morphologic reactions were observed in either the direct contact- or extract-exposed test wells. The EC₅₀¹ of the test material could not be determined, as viability quantitated with Neutral Red was not reduced regardless of the extract concentration (5, 50 or 100%). The test article passed both the direct contact and extract assays. Therefore, it was concluded that cured test article was not cytotoxic (2).

Two other cytotoxicity studies were conducted. In 1 of the studies, no cytopathic effects were reported for the test article or extracts (3). In the other study, the MEM extract did not produce a cytopathic effect, while a cytopathic effect was observed with both of the cottonseed oil extracts. The cytopathic effect noted with the cottonseed oil extract was attributed to the extraction medium (4).

¹ The concentration of extract needed to effect 50% of the cells.



USP Class V. The ability of the material to elicit an intracutaneous or systemic response according to the USP Class V procedure was determined in rabbits and mice, respectively. The extracts of the test article were prepared at a ratio of 3 cm² per ml (test article to volume of vehicle) with 0.9% Sodium Chloride for Injection (SCI), 5% Ethanol in SCI, Cottonseed Oil (CSO), and Polyethylene Glycol 400 (PEG-400) by heating at approximately 121°C for approximately 60 minutes in an autoclave. Blanks of each vehicle without test article were also extracted concurrently.

Intracutaneous Test. Six New Zealand White rabbits were divided into 2 test groups of 3 animals each. PEG extracts were diluted with SCI to 120 mg/ml for this test. All the animals in each group received 5 sequential 0.2 ml intracutaneous injections of 2 of the 4 sample extracts and their corresponding solvents, such that each extract was injected into 3 rabbits. Injections were done into back skin, on either side of the spinal column, at least 1" apart. The irritation response at each injected site was recorded at approximately 24, 48, and 72 hours after injection and compared to the score for the vehicle control. The intracutaneous injections of test article extracts in rabbits were no more irritating than controls for any solvent (5).

Systemic Toxicity Test. Four groups of 5 male mice were injected intravenously and 4 groups of 5 were injected intraperitoneally with extracts and blanks prepared in the solvents described above. Test article extracts and solvent blanks of PEG-400 were adjusted to 200 mg/ml whereas extracts and solvent blanks of other vehicles were used undiluted. The mice were observed for signs of toxicity immediately and at approximately 4, 24, 48, and 72 hours after injection. Body weights were taken before dosing and at approximately 24, 48 and 72 hours after injection. Slight weight losses (< 1.0 gram) were observed in 6 animals at 24 hours post-treatment (3 from the Ethanol/SCI extract, 1 each from the Ethanol/SCI control, PEG-400 extract, and CSO control groups); a weight loss of 2.6 grams was noted in 1 animal in the PEG-400 extract group. Slight weight losses (<1.0 gram) were also observed in 3 animals at 72 hours post-treatment (1 each from the SCI control, Ethanol/SCI extract, and PEG-400 control groups). None of these weight losses was considered noteworthy. Approximately 4 hours after dosing, 2 animals intraperitoneally injected with cottonseed oil control were observed to have wet fur in the urogenital region. The remaining animals did not exhibit any signs of acute systemic toxicity on Day 1, and no signs of systemic toxicity were noted at other observation points. Under the conditions of the study, none of the animals treated with the extracts of the test articles showed a biological reaction significantly greater than the animals treated with the solvent blanks. In addition, other acceptance criteria (body weight change, mortality, behavior change) for the test were met.



Based on these results, the test article was considered to have met the requirements of the USP Class V Biological Reactivity Test (6).

30-Day Implant. The cured test article was implanted into 2 groups of 3 New Zealand White rabbits, in the paravertebral muscles and abdominal subcutaneous tissues for periods of 7 and 30 days. The negative control material, USP Polyethylene Negative Control Plastic, was implanted in the same manner into corresponding sites on the opposite sides of each animal. The test article was received sterile in the form of rods 1 mm in diameter and 10-15 mm in length. The negative control article was cut into approximately 10 mm size pieces. Three rabbits were sacrificed for necropsy at 7 and 30 days post-implantation. The gross necropsy included observations of the thoracic and abdominal viscera and the collection and/or fixation of specific tissues for histological examination as needed. The implant sites were evaluated for infection, discoloration, necrosis, hemorrhage, and fibrous capsule formation. No remarkable test article-related findings were observed at any of the implant sites in any animal at either time point. In addition, the implantation sites were evaluated microscopically. After either 7 or 30 days of implantation, microscopic examination of the implants and surrounding tissue showed that the reaction to the test article was similar or less intense than the reaction to the control article. Furthermore, the tissue reactions at the test and control sites decreased at Day 30 compared to Day 7. Under the conditions of this study, the local tissue reaction elicited by the implantation of the test article was similar to the control article; therefore, the test article met the requirements of the Short-term Implant Test in rabbits (7).

REFERENCES

1. Dow Corning Internal Report – 2003-I0000-52951
2. Dow Corning Internal Report – 2003-I0000-52665
3. Dow Corning Internal Report – 1982-I0005-2159
4. Dow Corning Internal Report – 1982-I0005-2160
5. Dow Corning Internal Report – 2003-I0000-52730
6. Dow Corning Internal Report – 2003-I0000-52549
7. Dow Corning Internal Report – 2003-I0000-53032

Version Date: GBK/MDA 22 August 2005

Supercedes: 03 March 2004

In providing this information to you at this time, Dow does not undertake to do further testing, to monitor further developments or to provide further information to you. Dow does not make any warranty or representation, express or implied, with respect to the completeness or utility of the data or any other related



The Dow Chemical Company

information concerning the safety, efficacy and legal compliance of the end use application. To the full extent permitted by law, Dow disclaims any and all liability with respect to your use of, or reliance upon, any data, procedure, conclusions or opinion contained in the data provided or any related information.

Dow provides you with access to the data solely for your internal use and any submissions to governmental agencies by you, including for product (or chemical substance) registration purposes. Third parties may not use it or rely upon it. No third party obtains any right of any kind from the data or your use of it.

Page | 5 of 5